- (6) (a) J. B. Bapat, D. St. C. Black, R. F. C. Brown, and C. Ichlor, Aust. J. Chem., 25, 2445 (1972); (b) J. J. Tufariello and E. J. Trybulski, Chem. Commun., 720 (1973).
- (7) (a) L. A. Brooks and H. R. Snyder, "Organic Syntheses," Collect. Vol. III, Wiley, New York, N.Y., 1953, p 689; (b) M. A. Dmitriev, *Zh. Obshch. Khim.*, 931 (1964); *Chem. Abstr.*, 61, 5501b (1962).
 (8) E. R. Jones, E. Eglington, and W. C. Whiting, "Organic Syntheses," Collect. Vol. IV, Wiley, New York, N.Y. 1963, p.755.
- (9) H. Lindlar, *Helv. Chim. Acta*, 35, 466 (1952).
 (10) P. E. Eaton, G. F. Cooper, R. C. Johnson, and R. H. Mueller, *J. Org.*
- Chem., 37, 1947 (1972). (11) L. J. Loeffler, S. F. Britcher, and W. Baumgarten, J. Med. Chem., 13, 926 (1970).
- (12) G. H. Posner, C. E. Whitten, and P. E. McFarland, J. Amer. Chem. Soc., 94, 5106 (1972).
- (13) R. K. Crossland and K. L. Sevis, J. Org. Chem., 35, 3195 (1970).
- (14) N. Kornblum, H. C. Larson, R. K. Blackwood, D. D. Mooberry, E. P. Oliveto, and E. E. Graham, J. Amer. Chem. Soc., 78, 1497 (1956).

- (15) G. R. Delpierre and M. Lamchem, J. Chem. Soc., 386 (1960).
- (a) R. Huisgen, H. Houk, R. Grashley, and H. Seidl, *Chem. Ber.*, **101**, 2568 (1968); (b) R. Huisgen, H. Seidl, and I. Bruning, *ibid.*, **102**, 1102 (16)
- 1969)
- B. G. Murray and A. F. Turner, *J. Chem. Soc. C*, 1338 (1966).
 J. J. Tufariello, J. P. Tette, and E. J. Trybulski, unpublished results.
 N. L. Allinger, M. T. Tribble, M. A. Miller, and D. H. Wertz, *J. Amer. Chem. Soc.*, **93**, 1637 (1971).
- , K. R. Blanchard, and C. P. Woody, J. Amer. Chem. (20) P. v. R. Schlever
- Soc., 85, 1358 (1963). (21) L. I. Zakharkin, V. I. Stanko, and V. A. Brattsev, *Izv. Akad. Nauk SSSR*, Ser. Khim., 931 (1964). (22) D. Starr and R. M. Hixon, "Organic Syntheses," Collect. Vol. II, Wiley,
- (22) D. Starr and R. M. Hixon, "Organic Syntheses," Collect. Vol. II, Wile New York, N.Y., 1943, p 571.
 (23) S. Watson and J. Eastham, *J. Organometal. Chem.*, 9, 165 (1969).
 (24) J. Meinwald, J. Crandall, and W. E. Hymans, *Org. Syn.*, 45, 77 (1965).
 (25) G. B. Kauffman and L. A. Terer, *Inorg. Syn.*, 7, 9 (1963).
 (26) E. J. Corey and D. Beames, *J. Amer. Chem. Soc.*, 94, 7210 (1972).

Total Synthesis of β -Lactam Antibiotics. VI. 3-Arylcephalosporins¹

Raymond A. Firestone, Natalie S. Maciejewicz, and B. G. Christensen

Merck Sharp and Dohme Research Laboratories, Rahway, New Jersey 07065

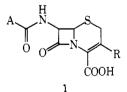
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The total synthesis of dl-3-phenyl-, 3-p-carbomethoxyphenyl-, and 3-(4-thiazolyl)-7 β -(2-thienyl)acetamidodecephalosporanic acids 12a-c is described.

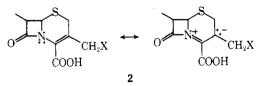
Cephalosporins 1 are a class of semisynthetic antibiotics that are being increasingly used because of their breadth of spectrum, potency, acid stability, and high degree of tolerance by man. In recent years an intensive worldwide effort has been made to obtain modified cephalosporins with improved properties.² Variations at the 3 position have been particularly fruitful, resulting in clinically useful drugs having such diverse substituents R as

-CH₂OAc, -CH₃, -CH₂Py⁺, -CH₂S-
$$\begin{pmatrix} N-N \\ S \end{pmatrix}$$
-CH₃

Many other modifications have also been reported, the vast majority of which have $-CH_2X$ as the 3 substituent,^{2,3} although R = H has also been reported.⁴



It is believed that resonance of type 2 plays a role in the bioactivity of cephalosporins, since electronegative groups X increase potency, and Δ^2 -cephalosporins are inactive.²



However, the activity of 3-methylcephems such as cephalexin (X = H) shows that X^- does not have to depart during the bioactive event. Furthermore, a theoretical study also concluded that during thiolation of cephalosporins at the β -lactam carbonyl, negative charge tends to accumulate at C-3 but that the CH₂-X bond does not break.⁵

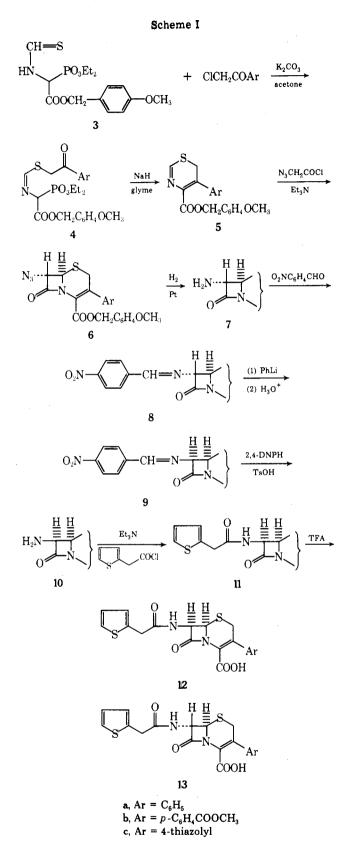
For these reasons it seemed worthwhile to prepare cephems bearing aromatic rings directly attached to C-3. At the inception of this project, there was no known way to do this by partial synthesis.⁶ However, the total synthesis recently developed in these laboratories⁷ has the capability of great variation in the 3 substituent, and this route (Scheme I) was therefore chosen for the preparation of 3-aryl cephems.

The alkylation of thioamide 3^7 with phenacyl chloride and cyclization of 4a to the thiazine 5a were best done sequentially, with isolation of 4a. When more than 1 equiv of K_2CO_3 was used with phenacyl chloride as had previously been done with 1-chloro-3-acetoxy-2-propanone,7 extensive decomposition occurred. Phenacyl bromide could be used in place of the chloride, but in addition to 4a it gave an isomer, presumably trans. Both isomers were stable to interconversion in refluxing CDCl₃.

Many conditions were tried for the cyclization of 4a to 5a, including K₂CO₃-acetone,⁷ KHCO₃-acetone, Et₃N-CHCl₃, PhLi-THF, NaH-THF, LDA-THF, and NaHglyme. Of these, the latter gave the cleanest product and was used in all subsequent work.

Cycloaddition of azidoacetyl chloride to 5a gave cephem **6a**, sometimes containing the Δ^2 isomer, which was formed from 6a with catalysis by triethylamine. The isomers were separable by chromatography, but with care the problem was avoidable altogether.

The stereochemistry of 6a was established as trans by the coupling constant of 1.5 Hz for H-6 and H-7, in accord with previous observations⁷ as well as our own subsequent examples. Since all naturally occurring cephalosporins and penicillins have cis stereochemistry, and trans compounds are inactive, it was necessary to epimerize the 7 substituent. This could not be done by simple equilibration because the trans isomers are generally the more stable ones, and so our procedure based upon steric approach control was used.⁸ Azidocephem 6a was reduced to amine 7a, which was converted to its Schiff base 8a with p-nitrobenzaldehyde. Formation of the 7 anion with phenyllithium, activation with DMF, and then acidification under kinetically controlled conditions, which occurs preferentially from the less hindered side, provided the cis Schiff base 9a with the natural configuration at C-7, along with recovered 8a, in a 2:1 ratio.



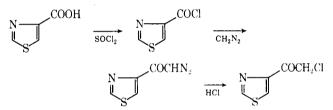
The steps $9a \rightarrow 12a$ were done in the usual manner,^{7,8} providing totally synthetic *dl*-3-phenyl cephem 12a. It contained some 7-epi isomer 13a carried through from the epimerization and some Δ^2 formed during thienylacetylation, but these, being inactive, did not interfere with the bioassay.⁹

With the expectation that an additional electron-withdrawing group would increase bioactivity, the *p*-carbomethoxyphenyl cephem 12b was also synthesized. The required *p*-carbomethoxyphenacyl chloride was prepared by the sequence below.

Our mp for the acid chloride, $50-55^{\circ}$, was different from that reported,¹⁰ 130°; so we prepared it by an alternate route to be secure.

The sequence of Scheme I was repeated, starting with 3 and p-carbomethoxyphenacyl chloride. The reactivity of the latter was much greater than that of phenacyl chloride, forming 4b in minutes instead of hours. The epimerization of 8b gave a normal:epi ratio (9b:8b) of only about 1:1, not separable at the 11b stage, so that the final product dl-12b contained also 13b in about a 1:1 ratio.⁹ Isomerization to Δ^2 was less than in the phenyl series, and in other respects the sequence was done in the same way as before.

To provide an example of a 3-heteroaryl cephem, the 4thiazolyl analog **12c** was made by the same route. The required chloromethyl 4-thiazolyl ketone was prepared from the acid as before.



The pathway of Scheme I was again followed, furnishing $dl-12c^9$ which was free of both Δ^2 isomer and 13c.

In all three syntheses, the yields of thiazines 5a-c were critically dependent on the purity of 3, which must be carefully chromatographed even though this does not change the ir or nmr spectra. We believe that crude 3, which is made from the amino compound by thioformylation in liquid H₂S, contains some form of active sulfur because sulfides of the type (ArCOCH₂)₂S (14) have been obtained from poor preparations of 5.

An unexpected transformation occurred on compounds 4a and 4c. On several days' standing, they were transformed into the formamide 15. We do not know the path-

$$\begin{array}{c} CHO \\ NH \longrightarrow PO_3Et_2 \\ COOCH_2C_6H_4OCH_3 \end{array}$$
15

way of this reaction, or the fate of the other fragment of the molecule.

A final point of interest concerns the conformation of the 3-aryl substituents. The phenyl and p-carbomethoxyphenyl groups have hydrogen atoms on both carbons adjacent to the point of attachment, which project in the plane of the ring and prevent it from becoming planar with the thiazine ring, in all compounds from **5a,b** to **12a,b**. On the other hand, the thiazole ring has only one such hydrogen, which is apparently not sufficient to prevent it from enjoying planarity, and thus conjugation, with the thiazine ring in **5c**-**12c**. We infer this from the nmr spectra on these grounds. In the **a** and **b** series relative to the **c** series, SCH₂, OCH₂Ar, and the two hydrogens in CH₂C₆H₄OCH₃ ortho to CH_2 appear *ca*. 0.2 ppm upfield. In the thiazolyl series, the latter two chemical shifts are about the same as those of the corresponding 3-alkyl compounds, showing that the protons in question are in the shielding cone of the phenyl rings in the **a** and **b** series.

The preparation of other totally synthetic β -lactam antibiotics will be reported in due course.

Experimental Section

Nmr spectra were taken on a Varian T-60; ir spectra were taken on a Perkin-Elmer Infracord.

Preparation of 4a–c. A mixture of 77 mg (0.2 mmol) of 3, 32 mg of phenacyl chloride (0.2 mmol), 29 mg of ground K_2CO_3 , and 2 ml of acetone was stirred 18 hr at 25° under N₂, filtered, and evaporated, affording 111 mg of crude **4a**, suitable for further reactions: 112% wt yield; ir (film) 5.72 (ester), 5.93 (COPh), 6.20, 6.28 μ (Ar, C=N); nmr (CDCl₃) δ 1.20 (t, J = 7 Hz, CH₂CH₃), 3.79 (s, OCH₃), 4.08 (d of q, $J_{HP} = 8$ Hz and J = 7 Hz, CH₂CH₃), 4.56 (s, SCH₂), 4.69 (d, $J_{HP} = 21$ Hz, CHP), 5.16 (s, OCH₂Ar), 6.9 (d, $J_{HP} = 4$ Hz, CH=N). When this reaction was done using 53 mg of phenacyl bromide, 37 mg of K₂CO₃, 100 mg of **3**, and 2 ml of acetone (0.267 mmol scale), nmr showed two products. From plc on silica gel with 4:1 CHCl₃-EtOAc was obtained 35 mg of **4a** and 34 mg of an isomer, ir similar to **4a**, nmr similar except for CH=N at δ 9.06 (d, $J_{HP} = 4$ Hz), mass spectrum 493. The nmr spectra of **4a** and its isomer did not change after brief boiling in CDCl₃.

Compound 4b was prepared by the same procedure but with only 25 min reaction time. The ir was similar to that of 4a with an added COOCH₃ at 5.78 μ , and the nmr was also similar, with an added COOCH₃ at δ 3.98.

Compound 4c was best prepared by a slightly modified procedure, using a threefold excess of K_2CO_3 and stirring 1 hr. After filtering, evaporating, and flushing with benzene, the crude 4c was ready for the next step. The ir and nmr spectra resembled those of 4a, lacking the C_6H_5 bands and with additional ones at δ 8.31 and 8.85 (m, thiazolyl).

Cyclization of 4a-c to 5a-c. Crude **4a**, prepared as above from 77 mg of **3**, was dissolved three times in glyme and evaporated and then treated 5 min in 1 ml of glyme with 1 equiv of NaH (10 mg of 50% NaH in oil, washed twice with hexane to remove oil) suspended in 1 ml of glyme. The reaction mixture was diluted with 10 ml of benzene, washed with water, dried with MgSO₄, filtered, and evaporated. Weight yields of crude **5a** generally ran >100%. Purification by chromatography was not successful, but crude **5a** was suitable for cycloaddition. The ir showed only ester carbonyl at 5.78 μ : nmr (CDCl₃) δ 3.58 (s, SCH₂), 3.76 (s, OCH₃), 4.93 (s, OCH₂), 6.71 (d, J = 9 Hz), 6.80 (d, J = 9 Hz, C₆H₄), 7.25 (s, C₆H₅), 8.30 (s, CH=N); mass spectrum 339.

Compound **5b** was made as above: ir 5.80 μ ; nmr (CDCl₃) δ 3.60 (s, SCH₂), 3.81 (s, OCH₃), 3.97 (s, COOCH₃), 4.98 (s, OCH₂), 6.78 (d, J = 9 Hz), 6.96 (d, J = 9 Hz, C₆H₄OCH₃), 7.27 (d, J = 8 Hz, C₆H₄COOCH₃), 8.40 (s, CH=N); mass spectrum 397. Crude yield >100%, 30% of crystalline **5b**, mp 144-6° from 1:1 benzene-cyclohexane.

Compound 5c was prepared similarly, ~100% crude yield, 39% crystalline, mp 134° from benzene-cyclohexane: ir 5.79 μ ; nmr (CDCl₃) δ 3.84 (s, OCH₃), 5.22 (s, OCH₂), 6.91 (d, J = 9 Hz), 7.27 (d, J = 9 Hz, C₆H₄), 8.43 (s, CH=N), 7.3 (d, J = 2 Hz), 8.75 (d, J = 2 Hz, thiazolyl); mass spectrum 346.

Cycloaddition of Azidoacetyl Chloride¹² to 5a-c Forming 6a-c. Crude 5a prepared from 0.25 mmol of 3, in 2 ml of CH₂Cl₂, was treated with 0.139 ml (1 mmol) of Et₃N. Then, under N₂, 0.033 ml of azidoacetyl chloride (0.37 mmol) in 2 ml of CH₂Cl₂ was added at 0° over 30 min. The mixture was allowed to warm to 25°, diluted with 30 ml of benzene, washed successively with pH 3 aqueous phosphate, water, and aqueous K₂HPO₃, dried with MgSO₄, filtered, and evaporated, leaving 106 mg of crude cephem 6a. This was chromatographed on 5 g of silica gel (E. Merck's) with 10:1 CHCl₃-EtOAc, providing 28.5 mg (27%) of pure 6a: ir (film) 4.73 (azide), 5.61 (β -lactam), 5.79 μ (ester); nmr (CDCl₃) δ 3.66 (s, SCH₂), 4.59, 4.70 (d's, J = 1.5 Hz, H-6 and -7), other peaks correct; mass spectrum 422, 394.

Crystalline **5b** was similarly treated with 4 equiv of Et₃N and 2 equiv of azidoacetyl chloride, affording 130 mg of crude (83%) and 29 mg of crystalline cephem **6b**. Chromatography of the crude gave 33 mg more **6b**, 40% total: ir like **6a**; mass spectra 480, 452; nmr (CDCl₃) δ 3.63 (s, SCH₂), 4.62, 4.70 (d's, J = 1.5 Hz, H-6 and -7), other peaks correct.

Crystalline 5c similarly gave, after chromatography, 28% of 6c: ir like 6a,b; mass spectrum 429, 401; nmr (CDCl_3) δ 3.81 (s, SCH₂ and OCH₃), 4.67, 4.73 (d's, J = 1.5 Hz, H-6 and -7), other peaks correct.

Caution! Vigorous explosions have been reported¹³ during the preparation of azidoacetyl chloride. Using a modified procedure¹⁴ we have had no trouble, but we urge the greatest caution in making and handling this compound.

Hydrogenation of 6a-c to 7a-c. Azidocephem 6a, 170 mg, was hydrogenated in 25 ml of benzene with 170 mg of PtO₂ for 45 min at 40 psi, filtered through a bed of Supercel and evaporated, affording 101 mg of 7a, 63%: ir (film) 5.66, 5.78 μ ; nmr (CDCl₃) δ 2.0 (m, NH₂), 3.59 (s, SCH₂), 4.16, 4.52 (d's, J = 1.5 Hz, H-6 and -7), other peaks correct. Also present was the Δ^2 isomer, nmr (CDCl₃) δ 4.26 (d, J = 1 Hz, H-7), 5.51 (m, H-6), 6.45 (d, J = 1.5 Hz, SCH=).

Likewise 6b was reduced to 7b, 67%: ir (film) 2.9, 5.63, 5.78 μ ; mass spectrum 454; nmr (CDCl₃) δ 2.3 (m, NH₂), 3.60 (s, SCH₂). 4.24, 4.58 (d's, J = 2 Hz, H-6 and -7), other peaks correct.

Similarly 6c gave 56% 7c: ir (film) 2.9, 5.64, 5.77 μ ; mass spectrum 403; nmr (CDCl₃) δ 2.25 (m, NH₂), 3.80 (s, SCH₂), 4.30, 4.60 (d's, J = 2 Hz, H-6 and -7), other peaks correct.

Preparation and Epimerization of Schiff Bases 8a-c to 9a-c. Compounds **7a-c**, *ca.* 2.5% in CH₂Cl₂, were treated 2 hr with 1 equiv of *p*-nitrobenzaldehyde in the presence of MgSO₄, filtered, and evaporated, providing 7α -Schiff bases **8a-c.** Excess of MgSO₄ should be avoided. Epimerization was done by the published procedure,⁸ giving mixtures of **8a-c** and the 7 β -Schiff bases **9a-c** in these ratios: **9a:8a**, 2:1; **9b:8b**, 1:1; **9c:8c**, 1:1. Epimerization was gauged by comparing in the nmr the CH=N peaks, δ 8.55 for 7α and δ 8.75 for 7β .

 $9a-c \rightarrow 10a-c \rightarrow 11a-c$. Deblocking of epimerized Schiff bases 9a-c and thienylacetylation of amines 10a-c were done as before.8 The crude amines were checked by ir only (similar to 7a-c) and carried forward without purification. Esters 11a-c were isolated by chromatography on silica gel (E. Merck, 70-230 mesh) in weight ratios of about 30:1, 40:1, and 50:1 respectively, eluting with 10:1 chloroform-ethyl acetate, and the single-spot products had R_{f} 's of about 0.3, 0.2, and 0.3, respectively, on the in the same system. Compound 11a had ir (film) 3.06 (NH), 5.62 (β -lactam), 5.77 (ester), 5.97 μ (amide); nmr (CDCl₃) δ 3.59 (s, SCH₂), 3.79 (s, OCH_3), 3.82 (s, $CH_2C=0$), 4.90 (s, OCH_2), 4.97 (d, J = 4 Hz, H-6), 5.87 (d of d, J = 4, 8 Hz, H-7), other peaks correct; mass spectrum 520, 340. Compound 11b had ir (film) 3.0, 5.61, 5.79, 5.95 μ; nmr (CDCl₃) § 3.58 (s), 3.78 (s), 3.84 (s), 3.93 (s, COOCH₃), 4.92 (s), 4.99 (d, J = 4 Hz), 5.86 (d of d, J = 4, 8 Hz), et al.; mass spectrum 578, 397. Compound 11c had ir (film) 3.04, 5.62, 5.80, 5.98 μ; nmr (CDCl₃) § 3.72 (s, SCH₂), 3.81 (s, OCH₃), 3.88 (s, CH₂C=O), 5.07 $(d, J = 4 Hz, H-6), 5.14 (s, OCH_2), 5.89 (d of d, J = 4, 8 Hz, H-7),$ et al.; mass spectrum 527, 347.

De-esterification of 11a-c to 12a-c. Samples of 20-25 mg of **11a-c** were taken up in 0.5 ml of anisole, cooled to 0°, and treated with 2.5 ml of trifluoroacetic acid for 4-5 min. Vacuum of <1 Torr was then applied while the sample warmed gradually to 30°; this removes first TFA, then anisole. Another 2.5 ml of anisole was added and pumped off to assure quantitative removal of TFA. The residue was taken up in a few ml of water containing excess NaHCO₃, washed 3 times with CH₂Cl₂, acidified with pH 2 phosphate, and extracted 3 times with CH₂Cl₂, acidified with pH 2 phosphate, and extracted 3 times with MgSO₄, filtered, and evaporated to obtain the free acids 12, or else re-extracted directly with water containing the calculated amount of NaHCO₃ and lyophilized to obtain 12a-c as sodium salts. Samples of 12a-c were treated with CH₂N₂ and sent for mass spectrum without purification.

Our general de-esterification procedure more recently is to treat one part ester with 2 parts anisole and 10 parts TFA for 2.0 min at 0° and work up as above. Sometimes good sodium salts are obtained simply by treating the residue with water containing the calculated amount of NaHCO₃, washing with CH_2Cl_2 , and lyophilizing. Yields vary from 60 to 95%.

Compound 12a had ir (film) 2.9-3.7, 5.60, 5.8, 6.0 μ ; nmr (Na salt, D₂O) δ 3.70, (s, SCH₂), 3.92 (s, CH₂C=O), 4.63 (HDO), 5.15 (d, J = 4 Hz, H-6), 5.67 (d, J = 4 Hz, H-7), et al.; also δ 6.50 (broad s, SCH= of Δ^2 isomer). High resolution mass spectrum of Me ester, 414.0713; calcd for C₂₀H₁₈N₂O₄S₂, 414.0707. Compound 12b had nmr (Na salt, D₂O) δ 3.68 (s), 3.88 (s, COOCH₃ and CH₂C=O), 4.68 (HDO), 5.16 and 5.67 (d's, J = 5 Hz), et al.; also δ 6.65 (m, SCH= of Δ^2 isomer). High resolution mass spectrum of Me ester, 472.0714; calcd for C₂₂H₂₀N₂O₆S₂ 472.076. Compound 12c's Me ester had mass spectrum 421, 241, 208, 181.

p-Carbomethoxybenzoyl Chloride. Following the published

procedure,¹⁰ dimethyl terephthalate was saponified to the monopotassium salt monoester and converted to the monoacid chloride monoester with SOCl₂. The crystalline product had good ir and nmr spectra, but melted at ca. 50° instead of the reported 130°. Therefore another method was also used. To a solution of 21.4 g (0.11 mol) of dimethyl terephthalate in 100 ml of MeOH and 100 ml of ether was slowly added a solution of 5.6 g (0.1 mol) of KOH in 100 ml of MeOH and 5 ml of H₂O. After being stirred for 18 hr the mixture had pH 7 and a white precipitate which was filtered, dissolved in H₂O, filtered to clarify, and acidified to pH 2 with HCl. A white precipitate of monomethyl terephthalate appeared which was filtered, washed, and dried, 11.4 g, mp 218-220° (reported¹⁵ 230°). One gram was refluxed 2 hr with 1.5 ml of SOCl₂ and evaporated, leaving a white powder, mp 50–55°, identical with the sample made by the first method: ir (Nujol) 5.63, 5.76 μ ; nmr (CDCl₃) § 3.98 (s, OCH₃), 8.15 (s, C₆H₄).

p-Carbomethoxydiazoacetophenone. Diazomethane was made by adding 27.2 g of Diazald in 165 ml of ether to 6.34 g of KOH in 10 ml of H₂O and 31.5 ml of EtOH. Over 1 hr, the CH₂N₂ solution was added to 6 g of p-carbomethoxybenzoyl chloride in 200 ml of ether at 0°. The volume was reduced to ca. 150 ml and the crystalline product was filtered and dried, 4.3 g, mp 98-101°: ir (CDCl₃) 4,74, 5,79, 6,16 μ; nmr (CDCl₃) δ 3.95 (s, COOCH₃), 5.93 (s, CHN_2), 7.81, 8.09 (d's, $J = 8 Hz, C_6H_4$).

p-Carbomethoxyphenacyl Chloride. The diazo compound, 4 was dissolved in 40 ml of AcOH and 4 ml of concentrated HCl. When effervescence ceased, 200 ml of H₂O was added, precipitating the product which was filtered, washed with water, and dried, 3.8 g, mp 144-7°: ir (CDCl₃) 5.79, 5.9 μ (sh); nmr (CDCl₃) δ 3.95 (s, $COOCH_3$), 4.70 (s, $COCH_2Cl$), 8.02, 8.07 (d's, J = 8 Hz, C_6H_4).

Thiazole-4-carbonyl Chloride. Thiazole-4-carboxylic acid, 0.97 g (0.01 mol), was refluxed with 10 ml of SOCl₂ for 90 min. The solution was evaporated and the residue flushed with benzene and recrystallized from 50 ml of hexane: mp 85-86.5°; ir (CDCl₃) 5.64 μ ; nmr (CDCl₃) δ 8.59, 9.01 (d's, J = 2 Hz).

4-Diazoacetylthiazole. The acid chloride, 4.01 g (0.027 mol), was added to 680 ml of 0.1 N CH_2N_2 in ether (0.068 mol) at 0°. The reaction took ca. 5 min and was stirred 30 min at 0° and 30 min at 25°. The solution was partly concentrated, filtered to clarify, and evaporated to dryness, leaving 4.4 g of yellow solid which was washed with petroleum ether; mp 80-82°. It was recrystallized from ether-petroleum ether and then from ether, giving 1.8 g of pure product: ir (CDCl₃) 4.72, 6.17 μ ; nmr (CDCl₃) δ 6.48 (s, $COCHN_2$), 8.27, 8.84 (d's, J = 2 Hz, thiazolyl).

4-Chloroacetylthiazole. The diazo compound, 3 g, was added at 0° to 50 ml of EtOH saturated with HCl. The mixture was stirred 5 min and evaporated to dryness. The solid was washed with a little ether, then stirred with pH 8 aqueous phosphate, and extracted 4 times with ether. The ether solution was dried with MgSO₄, filtered and evaporated, leaving 2.62 g of yellow solid, 83%; ir (CDCl₃) 5.85 µ; nmr (CDCl₃) δ 4.95 (s, COCH₂Cl), 8.40, 8.94 (m's, thiazolyl).

Sulfides 14a-c. From chromatography of cephems 6a-c and crystallization of thiazines 5b.c were obtained variable yields of 14a-c, depending on the quality of 3. Compound 14a had ir (film) 5.92 (sh), 5.97 μ ; mass spectrum 302 [(C₆H₅COCH₂)₂S₂], 270, 237, 165, 152, 119, 105, 77. Compound 14b had ir (CHCl₃) 5.80, 5.92, 5.97 μ; nmr (CDCl₃) δ 3.94 (s, COOCH₃), 3.97 (s, SCH₂), 7.98, 8.10 (d's, J = 8 Hz, C₆H₄); mass spectrum 386, 355, 353, 223, 178, 163, 135. Compound 14c, in mixtures with cephem 6c, had ir 5.93, 5.98 μ and a sample purified by tlc had mass spectrum 284, 158, 126, 112.84.

Formamide 15. On standing for several days, 4a and (more rapidly) 4c were transformed into 15: ir (film) 3.1-3.2, 5.74, 5.96 μ ; nmr (CDCl₃) δ 5.30 (d of d, $J_{\rm HP}$ = 22 Hz and J = 8 Hz, CHP), 8.20 (s, CH=O), et al.; mass spectrum 359, 331, 210, 195, 166, 138, 121.

Registry No.-3, 50917-87-8; 4a, 52539-73-8; 4b, 52539-74-9; **4c**, 52539-75-0; **5a**, 52539-76-1; **5b**, 52539-77-2; **5c**, 52539-78-3; **6a**, 52539-82-9; **6b**, 52539-83-0; **6c**, 52539-84-1; **7a**, 52539-85-2; **7a** Δ^2 isomer, 52539-79-4; 7b, 52539-86-3; 7c, 52539-87-4; 8a, 52539-88-5; 8b. 52539-89-6; 8c, 52539-90-9; 9a, 52539-91-0; 9b, 52539-92-1; 9c, 52539-93-2; 10a, 52539-94-3; 10b, 52539-95-4; 10c, 52585-05-4; 11a, 52539-96-5; 11b, 52539-97-6; 11c, 52539-98-7; 12a, 52539-99-8; 12a Na salt, 52540-00-8; 12a Δ^2 isomer Na salt, 52539-80-7; 12a Me ester, 52540-01-9; 12b, 52540-02-0; 12b Na salt, 52540-03-1; 12b Δ^2 isomer Na salt, 52539-81-8; 12b Me ester, 52540-04-2; 12c, 52540-05-3; 12c Na salt, 52540-06-4; 12c Me ester, 52540-07-5; 14a, 2461-80-5; 14b, 52540-19-9; 14c, 52540-20-2; 15, 52540-21-3; phenacyl chloride, 532-27-4; p- carbomethoxyphenacyl chloride, 52540-22-4; thiazole-4-carbonyl chloride, 52540-23-5; azedoacetyl chloride, 30426-58-5; p-carbomethoxybenzayl chloride, 7377-26-6; pcarbomethoxydiazoacetophenone, 22744-13-4; thiazole-4-carboxylic acid, 3973-08-8; 4-diazoacetylthiaziole, 52540-24-6; 4-chloroacetylthiazole, 52540-23-5.

References and Notes

- (1) Paper V: N. G. Steinberg, R. W. Ratcliffe, and B. G. Christensen. Tetrahedron Lett., in press.
- E. H. Flynn, Ed., "Cephalosporins and Penicillins," Academic Press, New York, N.Y., 1972. (2)
- J. H. C. Nayler, M. J. Pearson, and R. Southgate, Chem. Commun., 58 (3)(1973). R. Scartazzini and H. Bickel, *Helv. Chim. Acta*, **55**, 423 (1972)
- (4)
- W. C. Topp and B. G. Christensen, *J. Med. Chem.*, **17**, 342 (1974). Since that time we have prepared 3-heteroaryl cephems by partial synthesis but the method is not suitable for phenyl or substituted phenyl: J. (6) L. Fahey, R. A. Firestone, N. S. Maciejewicz, and B. G. Christensen, manuscript in preparation.
- R. W. Ratcliffe and B. G. Christensen, Tetrahedron Lett., 4645, 4649, 4653 (1973). (8) R. A. Firestone, N. S. Maciejewicz, R. W. Ratcliffe, and B. G. Christen-
- sen, *J. Org. Chem.*, **39**, 437 (1974). The bioactivities of these and other analogs prepared by total synthesis
- (9)will be reported elsewhere.
- (10) B. W. Hotten, *Ind. Eng. Chem.*, **49**, 1691 (1957). (11) We thank Dr. B. A. Arison for his invaluable assistance in the interpretation of nmr spectra. (12) A. K. Bose, B. Anjaneyulu, S. K. Bhattacharya, and M. S. Manhas, *Tet*-
- rahedron, 23, 4769 (1967).
- (13)Personal communication from Dr. J. L. Fahey.
- J. L. Fahey and R. W. Racliffe, manuscript in preparation.
 A. Baeyer, *Justus Liebigs Ann. Chem.*, **245**, 103 (1888).